



Production Assistance for Cellular Therapies  
National Heart Lung and Blood Program



National Heart, Lung,  
and Blood Institute

# Minimum Characterization Criteria for Clinical Grade iPSC Cell Banks and Final Products

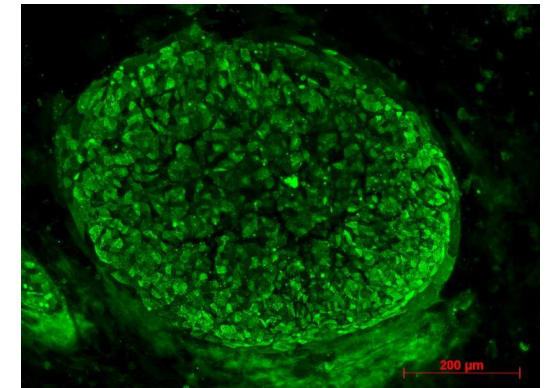
**Stakeholder Association: NHLBI, PACT**

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# Induced pluripotent stem cells (iPSCs) - Need to establish guidelines for minimum characterization criteria

- A crucial problem in both the analysis of many human diseases and the development of effective therapies to treat disease is the incomplete understanding of the role played by human genetic variation in their development.
- Guidelines need to be established in the following area:
  - Donor qualification CFR 1271
    - Genetic testing
  - iPSC cell bank testing
  - iPSC final product testing



# Challenges

- iPSC derivation (e.g., safety and efficiency of the reprogramming method, donor-to-donor variability, and choice of starting materials)
- iPSC challenges (e.g., development of a cell culture system for iPSC generation and expansion, cell sensitivity and robustness, and cryopreservation and revival)
- Safety and QC challenges (e.g., normal karyotype, residual plasmid clearance, in-process controls to evaluate iPSC quality, critical attributes of the final products, and standard safety concerns such as sterility).

# Pros, Cons and Unknowns to iPSC Technology

- Pros:
  - Eliminate ethical issues
  - Donor's clinical phenotype is often known when working with iPSCs.
- Cons:
  - Cells would still have genetic defects
  - One of the pluripotency genes is a cancer gene
  - Viruses might insert genes in places we don't want them (causing mutation)
  - iPSC treatments will likely require donor's cells to undergo genetic alterations. Will it be acceptable to people that their cells have been modified?
- Unknowns:
  - could genetic defects affect recipients
  - could reprogramming alter or carry genetic defects

# Identity - Cellular Biological Properties & Heterogeneity and Pluripotency

- Morphology
- Growth properties
- Stem cell markers: iPSCs expressed cell surface antigenic markers expressed on ESCs. Human iPSCs expressed the markers specific to hESC, including SSEA-3, SSEA-4, TRA-1-60, TRA-1-81, TRA-2-49/6E, and Nanog.
- Stem Cell Genes: Oct-3/4, Sox2, Nanog, GDF3, REX1, FGF4, ESG1, DPPA2, DPPA4, and hTERT.

# Donor Qualification

- Allogeneic
  - Donor qualification 21 CFR 1271
  - How does donor age influence the reprogramming process and iPSC functionality?
  - Perform **genetic testing**

# Differentiation Potential

- Gene expression
  - Endoderm
  - Ectoderm
  - Mesoderm

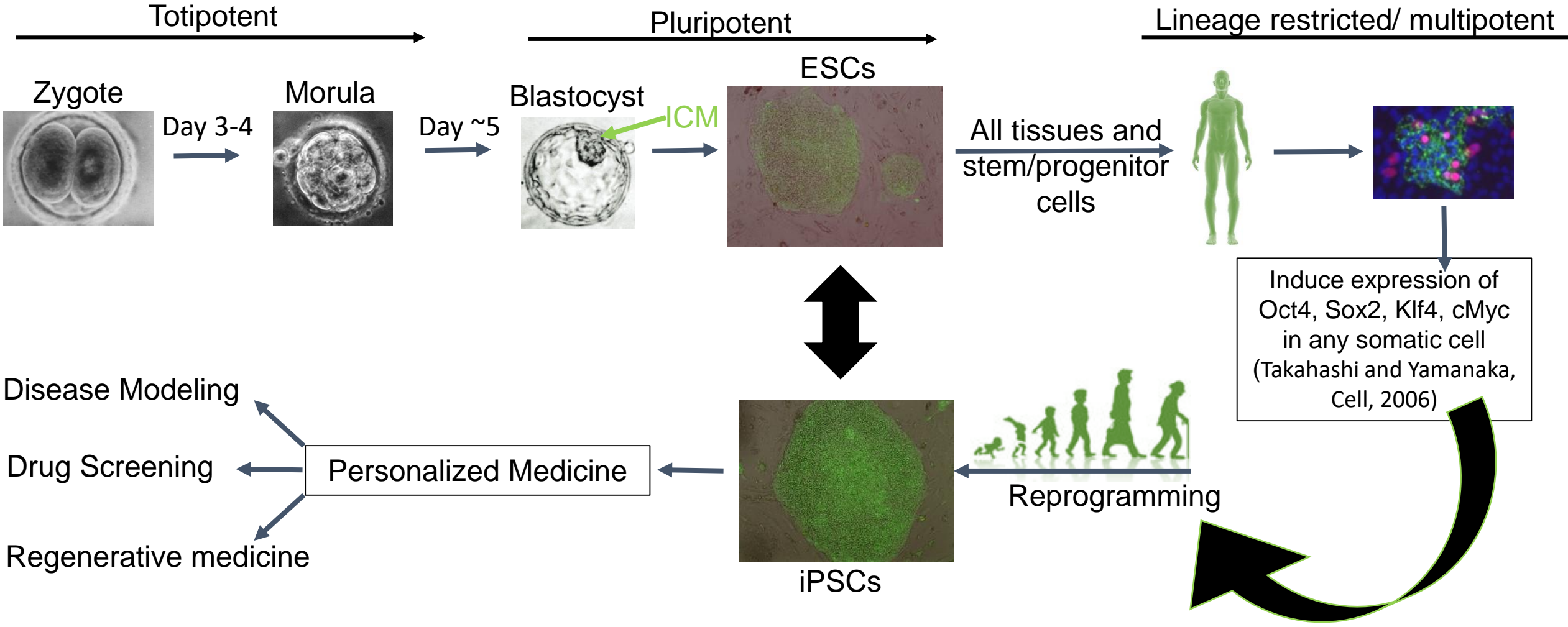
In vitro differentiation assay. (a) Confirmation of the differentiation potential of iPSC through spontaneous differentiation and formation of Embryoid body. (b) Analyze differentiation for the presentation of the markers of cells of the **3 germ layers**, by qPCR of the Embryoid body.

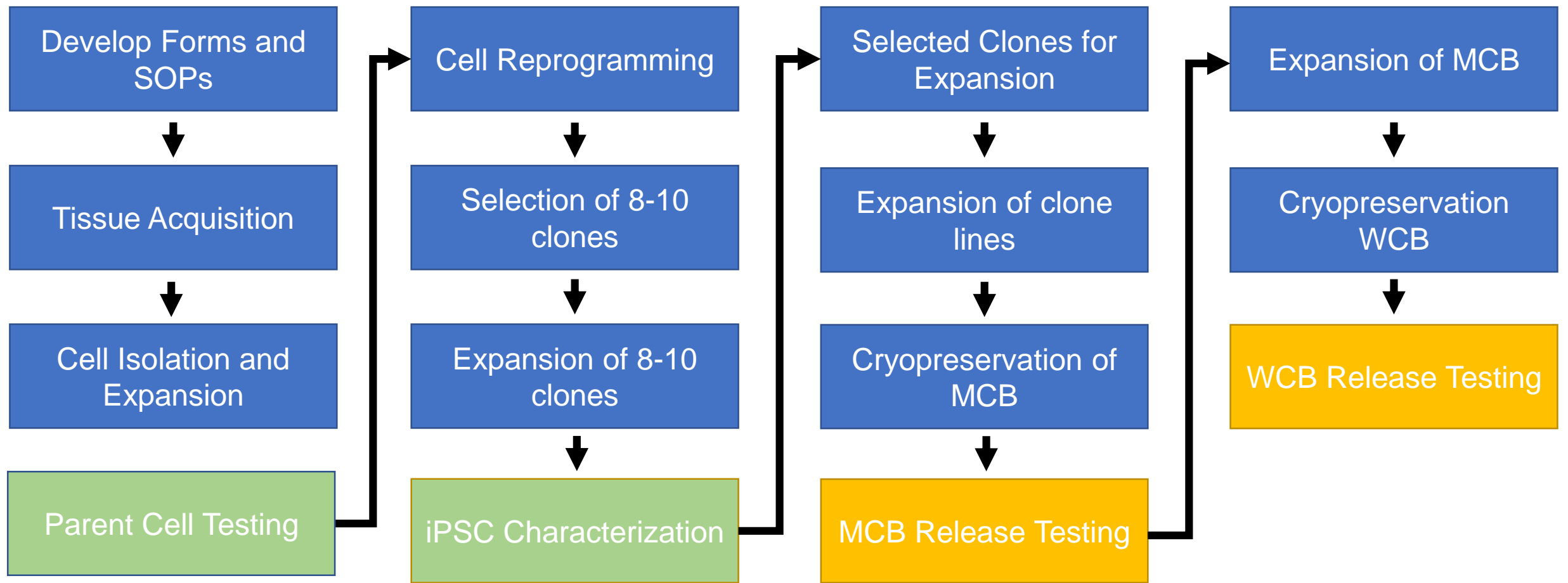
# Genome Integrity

- iPSC genome can reproduce that of the cell from which they are derived.
- iPSCs can potentially develop genetic abnormalities during reprogramming or prolonged cell culture.
- iPSC genome integrity must be routinely monitored by **karyotyping**



# Induced Pluripotent Stem Cells (iPSCs)





Yellow= MCB and WCB End Product Testing  
 Green= In progress Testing

# Parent Cell Testing

Objective	ASSAY	Acceptance Results
Sterility	Immersion Sterility tests Bacteriostasis and Fungistasis (B/F)	No organisms Pass
Mycoplasma Testing	Points to consider mycoplasma detection	Negative
Endotoxin	Kinetic chromogenic LAL	< 0.5EU/mL
Cell Viability	Trypan Blue	>75%
Genetic Stability	Karyotype Analysis	46, XY
	STR Genotyping	Retain STR profile of parent population for later comparison

# iPSCs Characterization

Objective	ASSAY	Acceptance Results
<b>Sterility</b>	Immersion Sterility tests Bacteriostasis and Fungistasis (B/F)	No organisms Pass
<b>Mycoplasma Testing</b>	Points to consider mycoplasma detection	Negative
<b>Endotoxin</b>	Kinetic chromogenic LAL	< 0.5EU/mL
<b>Cell Viability</b>	Trypan Blue	>75%
<b>Reprogramming Vehicle Residual Testing</b>	Depends on the method used for reprogramming	Negative
<b>Genetic Stability</b>	Karyotype Analysis	46, XY
	STR Genotyping	STR profile of starting population and iPSC lines are identical
<b>Pluripotency Phenotype</b>	Flow cytometry	SSEA4 > 70% Tra-1-60 > 70% Tra-1-81 > 70%
	Immunostaining	NANOG > 70%; OCT3/4 > 70% SSEA4 > 70% Tra-1-60 > 70%; Tra-1-81 > 70%
	Embryoid body formation	Detect of at least one marker per germ layer

# MCB and WCB Release Testing

Objective	ASSAY	Acceptance Results
<b>Sterility</b>	Immersion Sterility tests and Bacteriostasis and Fungistasis (B/F)	No organisms
<b>Mycoplasma Testing</b>	Points to consider mycoplasma detection	Negative
<b>Endotoxin</b>	Kinetic chromogenic LAL	< 0.5EU/mL
<b>Cell Viability</b>	Trypan Blue	>75%
<b>Genetic Stability</b>	Karyotype Analysis	46, XY
	STR Genotyping	STR profile of starting population and iPSC lines are identical
<b>Viral Contaminant</b>	In vitro Assay for Adventitious Virus Contaminant	Negative
	Fluorescent Product Enhanced reverse Transcriptase (FPERT) method	Negative
	Transmission Electron Microcopy (TEM)	No viral particles
<b>Pluripotency Phenotype</b>	Flow cytometry	SSEA4 > 70% Tra-1-60 > 70% Tra-1-81 > 70%
	Immunostaining	NANOG > 70%; OCT3/4 > 70% SSEA4 > 70% Tra-1-60 > 70%; Tra-1-81 > 70%
	Embryoid body formation	Detect of at least one marker per germ layer

# Future Direction

Explore the possibility of using single-cell RNA-seq to identify unique subpopulations that are especially suited for a patient-specific therapy.

# Conclusion

Using autologous or allogenic iPSCs has advantages and disadvantages, and the choice of appropriate strategy may vary depending on the intended use. Additionally, there remain many factors that affect establishing transplantation therapy using iPSCs. To avoid tumorigenesis and establish effective differentiation into the intended cells, further investigation is needed to clarify which iPSC line is the most suitable and how these lines can be best selected.